

Population estimation of a cryptic moss frog using acoustic spatially explicit capture recapture

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Abstract—Cryptic amphibians pose a problem for conservation managers, as they are difficult to find to assess initial populations and monitor changes during potentially threatening processes. The small Rough Moss Frog, *Arthroleptella rugosa*, occurs in seepages on a single unprotected mountain in South Africa's fire prone, biodiverse fynbos biome. The area is heavily impacted by invasive plants, which dry seepages and increase the frequency and intensity of fires, leading to the assessment of this species as Critically Endangered. We aimed to test the efficacy of acoustic spatially explicit capture recapture (aSCR) to estimate the entire population size of calling adult *A. rugosa* and assess the impacts of invasive plants. Our aSCR estimates suggest that the *A. rugosa* population is at ~2000 individuals, which is more than five times larger than previously estimated using aural calling surveys on the mountain, despite an intense fire over the entire area three years earlier that reduced the calling population to a few tens of individuals. Our vegetation surveys suggest that ongoing removal of invasive plants from the mountain is successful in areas occupied by *A. rugosa*, but that adjacent areas invaded by pines (*Pinus pinaster*) and hakea (*Hakea sericea*) have a negative impact on calling density. The private–public conservancy partnership on Klein Swartberg Mountain is conserving this frog, but ongoing management and monitoring are required to ensure conservation in the future.

Keywords— acoustic survey; fire; invasive plants; off-reserve matrix; population estimation; spatial capture recapture

Introduction

Globally, threatened and range restricted species are of high conservation concern (Gaston and Fuller 2009). Species with a restricted geographic range are often habitat specialists and weak dispersers, and thus face an increased risk of extinction due to threatening processes such as climate change, human development, and the spread of invasive species and diseases (e.g., amphibians [Sodhi *et al.* 2008, Cooper *et al.* 2008, Harper *et al.* 2022], mammals [Cardillo *et al.* 2008], birds [Lee and Jetz 2011], and plants [Casazza *et al.* 2014]). The rapid and increasing changes to habitat suitability and fragmentation are likely to, and in some cases already have, exceed restricted-range species' migration capabilities (Pearson 2006, Casazza *et al.* 2014). In some countries, species listed as threatened by the International Union for Conservation of Nature (IUCN) are often afforded protection that requires impact mitigation through the protection, restoration, or creation of habitat (Rodrigues *et al.* 2006), but in others no degree of threatened status affords any tangible protection. Despite a global bias towards protecting range-restricted and threatened species, there is still a tendency for threatened species to be poorly represented in protected area networks across the globe (Rodrigues *et al.* 2004, Nori *et al.* 2015). Amphibians are the least represented taxon, with almost a quarter (1,535 species) of all known extant amphibian species (~ 6,500) remaining unrepresented in protected areas (Venter *et al.* 2014, Nori *et al.* 2015), and most of these unrepresented

species occur in only one site (Ricketts *et al.* 2005). Adding to the complexity of amphibian conservation is our general lack of understanding of the conservation status of those species or communities typically found within the off-reserve matrix (*i.e.*, those typically excluded from reserve networks). Funding in the off-reserve matrix is often far more limited compared to resources that are linked to specific protected area management activities. Furthermore, monitoring rare and range-restricted species can require large sampling efforts that can be difficult and impractical.

In South Africa, the most threatened native amphibian species are concentrated within the Cape Floristic Region, many of which are affected by agriculture (50%), invasive species (37.1%) and habitat change and loss (25.9%; Stuart *et al.* 2004, Angulo *et al.* 2011, Mokhatla *et al.* 2012, Harper *et al.* 2022). Within this 8.77-million-hectare (ha) region, protected areas cover over 2.3 million ha (26.6%) of land, and 96,557 ha (1.1%) of land is comprised of privately-owned stewardship areas (Rouget *et al.* 2014). In these privately-owned areas there is a recognized need to integrate monitoring for improved conservation in an adaptive management framework (Rouget *et al.* 2014). Further, over half of the seepage habitat that many amphibian species rely on is Critically Endangered, with only 10% considered to be well protected (Driver *et al.* 2012). Annually, invasive plant species alone cause a loss of up to 87 million m³ per year of mean annual surface water runoff across South Afri-

ca (Wilson *et al.* 2014), which can be a potentially fatal change for native amphibians. With only 12.5% of the natural threatened habitat area remaining in the Western Cape, the control of invasive species has been prioritized and is estimated to exceed an annual cost of \$23.5 million USD (Wilson *et al.* 2014).

Many of the amphibian species within South Africa are poorly studied, and their population responses to these threats remain poorly understood (Measey 2011, Measey *et al.* 2019). *Arthroleptella rugosa*, a Critically Endangered moss frog (IUCN & SA-FRoG 2016), is restricted to a single mountain in the off-reserve matrix of the Western Cape. It is currently threatened by non-native invasive plants (*Pinus pinaster* and *Hakea sericea*), which degrade and dry their seepage habitat, and by fires in the invaded fuel-laden vegetation that are more severe than those with which it evolved (Turner and Channing 2008, IUCN & SA-FRoG 2016). These synergistic threats are thought to severely impact *A. rugosa* populations. As *A. rugosa* occurs exclusively on private land, it is dependent on the continued conservation efforts of the Klein Swartberg Conservancy, a collective of private landowners who designated the entire 14,857-ha mountain as a conservation area. CapeNature, the provincial nature authority, undertakes annual monitoring on the mountain to assess specific populations within the range of *A. rugosa*. Using aural calling surveys of the mountain (Dorcas *et al.* 2009), these survey techniques estimated the known populations to be around 400 adults (Turner and Channing 2008). This figure was doubled to account for non-calling females and rounded up to account for imperfect detection of males during the brief survey, resulting in an estimate for the species of 1,000 individuals in the IUCN Red List assessment (IUCN & SA-FRoG 2016). An intense fire over the entire mountain in January 2012 was followed by further aural acoustic surveys that suggested the population was heavily impacted, with only a few tens of individuals calling.

Conventional amphibian population monitoring is often carried out through visual surveys, trapping methods (capture-mark-recapture [CMR]), or through acoustic surveys of calling males (Driscoll 1998, Dorcas *et al.* 2009, Marsh *et al.* 2017). Although these methods are commonly used and often largely successful, some can be impractical for cryptic or small-bodied species, particularly for those with small population sizes, such as *A. rugosa*. An acoustic approach, vocally ‘capturing’ individuals, is less invasive and can be well suited to these hard-to-find species. Calling surveys provide rapid data that can be used to identify unnamed species, map distributions for occupancy models, and deduce qualitative count data (Dorcas *et al.* 2009, Marsh *et al.* 2017). Aural calling surveys, however, can be subject to imperfect detection, misidentification, and inconsistencies if conducted by inadequately trained observers (Dorcas *et al.* 2009). Observation errors such as omission (failure to detect a species that is present) and commission (incorrectly ‘detecting’ an absent species; Parris *et al.* 1999, Tyre *et al.* 2003) make it difficult to reliably measure species occurrence or population size, which can significantly affect population models and derived population estimates (Rogers *et al.* 2013). The use of automated recording systems eliminates the dependence on real-time processing by skilled

observers and allows researchers to collect permanent records of data, which can be examined and verified later (Dorcas *et al.* 2009). However, the applications of estimates attained using both aural calling surveys and traditional recording systems are limited, as the sampling area cannot be clearly defined and thus no density or actual population size estimates can be deduced (Stevens *et al.* 2002, De Solla *et al.* 2006).

Spatial capture-recapture (SCR; Efford 2004, Borchers 2012, Borchers and Fewster 2016) is a more recently developed method that combines capture-mark-recapture (CMR) and distance sampling methods (Buckland *et al.* 2001, Stevenson *et al.* 2015). While spatial capture-recapture was originally designed as a physical trapping method, it has since been adapted to use acoustic techniques to efficiently and non-invasively collect and analyze large volumes of acoustic data (Dawson and Efford 2009, Efford *et al.* 2009, Marques *et al.* 2012, Stevenson *et al.* 2015, Measey *et al.* 2017, Stevenson *et al.* 2021). This adapted technique, ‘acoustic spatial capture-recapture’ (aSCR), uses an array of fixed microphones to estimate the population density of vocalizing individuals and could be a suitable replacement for traditional survey methods typically used for visually cryptic, threatened and vocally distinct anuran species (Efford *et al.* 2009, Stevenson *et al.* 2015). While aSCR relies solely on calling individuals to determine population densities, these density estimates have been shown to be comparable to CMR estimates and thus can be reliable indicators of population trends and sizes (*e.g.*, Meuche and Grafe 2005). Currently, aSCR is the only known acoustic density estimation method that can also generate confidence intervals in a statistically rigorous manner (Measey *et al.* 2017).

We implement this non-intrusive aSCR technique to efficiently estimate (1) the total adult male *A. rugosa* population across their range and (2) *A. rugosa* adult male population densities in sites currently monitored and managed by conservation managers and private landowners. We assess the recovery of the population after the intense fire in 2012, and we use vegetation surveys in and adjacent to our recording areas to assess the efficacy of ongoing conservation efforts to control invasive plant populations. Population data will be used to assess the current *A. rugosa* population and conservation status, and direct ongoing and future efforts of private landowners and conservation managers towards areas of high importance for this species.

Methods

Study species—*Arthroleptella* is a genus of moss frogs within the speciose family Pyxicephalidae that are endemic to sub-Saharan Africa (Van der Meijden *et al.* 2011, Rebelo and Measey 2019). *Arthroleptella rugosa* is the most threatened and range-restricted species within this genus, occurring exclusively on private land on the Klein Swartberg Mountain (see Fig. 1). This species is typically found within, or near, seepages or wetland flats that have moist soil and adequate vegetation to provide protection from strong winds and high temperatures. Adults are small (mean body length of 13 mm; Turner and Channing 2008) with a dark brown appearance (Fig. 1). Their size and cryptic coloration makes finding individuals difficult; however, like other *Arthroleptella* species, *A. rugosa* males are

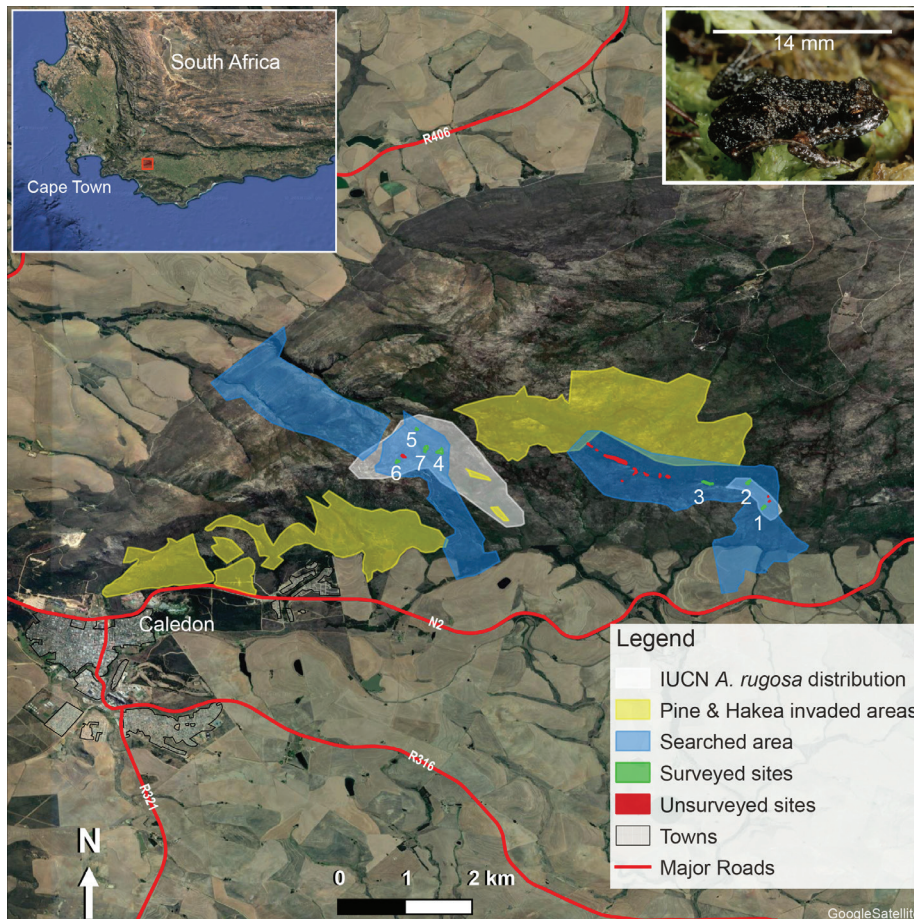


Fig. 1. Map of the study area, with inset (top left) showing the position of the site in the southwest of South Africa, and (top right) an *Arthroleptella rugosa* male with SVL 14 mm. The map demonstrates the search effort (blue polygons), identified areas of *Pinus pinaster* and *Hakea sericea* invasion (yellow polygons), surveyed habitat patches (green polygons), unsurveyed habitat patches (red polygons), and the IUCN listed *A. rugosa* habitat (grey polygons). Survey areas 1–3 are within the Eastern portion of the distribution and survey areas 4–7 are located within the Western portion.

easily distinguishable and detectable by their advertisement calls (Turner and Channing 2017).

Arthroleptella males are known to call from within seepage areas during an extended period throughout the austral winter; typically, May to November (Measey *et al.* 2017). Males exhibit three distinct vocalizations: a chirp-like advertisement call, a low frequency aggressive call, and a chuckling call unique to this species (Turner and Channing 2008). Calling behavior is largely unknown; however, during the breeding season, males are known to frequently vocalize advertisement calls, as these are presumably used for attracting female mating partners and defining territorial boundaries to conspecifics (Turner and Channing 2008, Angus *et al.* 2023). Advertisement calls have been shown to be the most frequently vocalized calls and are typically easy to capture in recordings (Kohler *et al.* 2017).

The entire population of *A. rugosa* occurs within a 230-ha area, and has previously been estimated as 400 adults, although this is expected to be in decline due to the ongoing plant invasion of *Pinus pinaster* and *Hakea sericea* (Turner and Channing 2008, SA-FRoG & IUCN 2016). A fire in January 2012 burned the entire mountain (Turner 2012, Measey *et al.* 2019,

Angus *et al.* 2023), and the following aural acoustic survey suggested the *A. rugosa* population was heavily impacted, with only a few tens of individuals calling in two areas of the eastern portion of the range (17 May 2012; AT and JM, *pers. obs.*). The increasing presence of invasive plants reduces the availability of water and increases the frequency and intensity of fires that naturally cycle in this area (Le Maitre *et al.* 2002, Measey 2011). Consequently, a focused effort to remove invasive vegetation from the mountain was started in 2012 under the auspices of CapeNature and supported by the U.S. Fish and Wildlife Service. Due to the level of threat currently facing this species, its conservation status, and the limited scientific literature available on the ecology of this species, *A. rugosa* has been identified as a high priority species requiring further research (Measey 2011).

Study area—The study area was located on private lands on the Klein Swartberg Mountain near Caledon, South Africa (34°12'S 19°32'E; Fig. 1). The area is characterized by Overberg Sandstone and Western Coastal Shale Band Vegetation Fynbos (Mucina *et al.* 2012) and hosts the entire *A. rugosa* population. Structurally, overstory proteoid and asteraceous

shrubs dominate the Klein Swartberg Mountain, creating an evergreen, fire-prone mosaic of open, mid-dense and closed vegetation coverage (Cowling *et al.* 1997, Mucina *et al.* 2012).

Arthroleptella rugosa habitat patches were identified based on previous records (Fig. 1), habitat type, and thorough area searches. Searches were conducted in areas of recognized suitable habitat and consisted of two researchers walking approximately 50 m apart listening for *A. rugosa* calls (blue polygons; Fig. 1). Populations occur in small habitat patches sporadically dispersed along the main mountain ridge (green and red polygons; Fig. 1). The area of identified *A. rugosa* patches, and thus the actual spatial extent of their distribution, was determined to facilitate an accurate calculation of the total population size using estimated densities.

Acoustic surveys—Seven sites ranging from the eastern to the western side of the *A. rugosa* distribution were selected based on logistic feasibility for acoustic surveying and were sampled from July through to September 2015 to coincide with the breeding season and associated peak calling times (green polygons; Fig. 1). Each site was surveyed three times to account for imperfect

detections associated with fluctuating environmental conditions or survey time.

For each survey, six Audio-Technica AT8004 Handheld Omni-directional Dynamic Microphones were connected to a Tascam DR-680 6-Track Portable Audio Recorder and set to record to six independent but synchronous tracks with a resolution of 24-bit and a recording frequency of 48 kHz. Microphones were attached to 1-m wooden dowels that were inserted into plastic tubing arranged in an array positioned in the center of the habitat patch of calling *A. rugosa* males. The plastic tubing remained at each site between surveys to keep microphone positions constant throughout the survey period. Straight-line distances between all microphone pairs were measured to the nearest centimeter and the GPS locations (Garmin GPSMap64) of each microphone were recorded. Once recording commenced, the immediate survey area was vacated, and *A. rugosa* males were recorded for 40 minutes. Surveys were conducted exclusively in good weather conditions (*i.e.*, no rain, minimal wind) to maximize individual call detectability (see Measey *et al.* 2017 for more details on methodology).

Data pre-processing—Raw acoustic recordings were processed to identify *A. rugosa* detections using the open-source software for Passive Acoustic Monitoring, PAMGuard (Gillespie *et al.* 2009). The PAMGuard *Click Detector* was configured to detect the *A. rugosa* advertisement call using characteristics identified in Turner and Channing (2008). For each detection, the start time, relative amplitude, and identification number of the microphone that made the detection were recorded. The start time for each call was recorded with an accuracy of 2.083×10^{-5} seconds.

Acoustic spatially explicit capture-recapture (aSCR)—The capture histories (call start time, relative amplitude, and detection information) obtained during pre-processing were used to determine the *A. rugosa* calling male density (frogs per hectare) using acoustic spatially-explicit capture-recapture (aSCR), a novel methodology outlined in Stevenson *et al.* (2015) and further applied in Measey *et al.* (2017). In processing the PAMGuard detection outputs, the first five minutes of each recording were omitted from the data to account for disturbance to the survey area caused by the presence of researchers. The detections (and non-detections) of calls across the array, and associated amplitudes and start times for each detection, facilitated the approximation of frog call locations. Detection probability functions were estimated using call and microphone locations (collected in the field but corrected according to the distances between microphone pairs for precision; see Measey *et al.* 2017) and used to determine the probability that a call emitted from any location in the survey area was detected by at least one microphone. From this, the proportion of calls detected across the survey area and the estimated area in which these calls were made over the survey period (Effective Survey Area; ESA) were calculated. The call density was estimated by dividing the total number of calls by the ESA and the survey length.

The use of aSCR requires an understanding of the target species vocal behaviors (Marques *et al.* 2013). Specifically, ac-

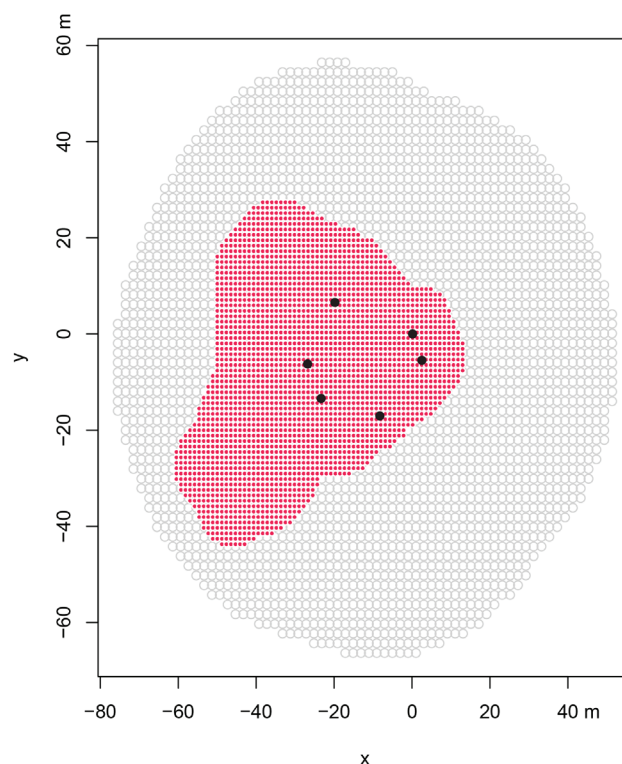


Fig. 2. Graphical representation of the previously used 40-meter buffer of assumed frog presence (gray circles) and an example of the input used to delineate the extent based on suitable habitat (red circles). Black dots represent exact placement of microphones in the field, with x and y coordinates in meters.

curate vocalization call rates estimates are necessary to precisely estimate calling animal density. During each acoustic survey, individual males were located and recorded for 20 minutes using a directional hand-held recorder (Olympus LS-10) to attain samples of *A. rugosa* advertisement calling ($n = 22$). Recordings of individuals were manually annotated in the open source software Audacity (see <http://audacityteam.org/>) to determine the number of *A. rugosa* advertisement calls made per minute.

Incorporation of population boundaries—In previous aSCR studies, each survey area has been defined by a buffer around the microphone array (Stevenson *et al.* 2015, Measey *et al.* 2017; *e.g.*, gray circles in Fig. 2). This assumes that calling males are present throughout this extent. Knowing, however, that the distribution of *A. rugosa* males does not conform to this spatial arrangement, this study delineated the actual patch area by using the path function on a Garmin GPSMap64 during area searches (*e.g.*, red circles in Fig. 2). This masking process improves the precision of call location estimates, and thus also the detection function and density estimates.

Population estimates—The R package *ascr* (Stevenson and Borchers 2015) was used to determine the detection function estimates and the call density. Modeling was computationally intensive, so 10 2-minute subsamples were analyzed for each recording, giving a total of 20 minutes for analysis.



The density estimate for each surveyed patch was multiplied by the relevant patch area (in ha) to obtain site-specific population size estimates. The population sizes for the 11 unsurveyed habitat patches were determined by multiplying the mean density from the seven surveyed sites by the total area of the unsurveyed *A. rugosa* habitat (2.42 ha). The total *A. rugosa* adult population size across the Klein Swartberg Mountain was a summed estimate of population sizes from all surveyed and unsurveyed populations (multiplied by two to account for non-vocalizing females, assuming a sex ratio of 1:1). We acknowledge that bias in this estimation of the total population is inherent as we were not able to randomly sample *A. rugosa* habitat patches.

Quantifying uncertainty in density estimates—A parametric bootstrap included in the *aSCR* package was used to estimate the uncertainty of the density estimates calculated. Call rate data were included in the bootstrap simulation to account for the dependence between call locations and to attain the calling male densities with standard errors for each survey area. The bootstrap procedure was run for 500–1,000 iterations. To correct for inconsistent results between bootstrap-procedure repetitions, a secondary bootstrap was run concurrently for 500 iterations to quantify the *Monte Carlo Error* (MCE) associated with the density and standard error estimates generated. The number of bootstrap iterations for each analysis was increased until the Monte-Carlo error was below 0.05, where the MCEs for all parameters are not more than 5% over their standard error. This ensured that the number of bootstrapping iterations were sufficient to provide accurate parameter estimates.

The total population size estimate was calculated as a sum of the estimated populations within surveyed areas and the estimated population within the unsurveyed area. The population estimate for each surveyed area was calculated using the average density and associated standard error for that site from aSCR estimates, whereas the *A. rugosa* populations within unsurveyed habitats were calculated using the reported average density of all sites surveyed, and the standard error of this total population was calculated from the difference of all density estimates, ignoring the error generated for each individual site.

Habitat assessments—Habitat assessments were carried out in each survey area to (1) identify the typical features of *A. rugosa* habitat and (2) assess *Pinus pinaster* and *Hakea sericea* invasion. Ten 1-m² quadrats were randomly placed throughout each survey area. For each quadrat, the percentage of ground cover (*i.e.*, grasses and restios) within the quadrat was estimated and the heights of at least three plants occurring within the quadrat were recorded. Where invasive vegetation was present within a survey area, the number and height of plants were also recorded. Additionally, the level of invasive vegetation (adult and juvenile plants) present surrounding the survey area was categorized into low (< 5 plants), medium (5–15 plants) and high (> 15 plants).

Statistical analysis—Summary statistics were calculated to identify typical *A. rugosa* habitat characteristics. A linear model

was performed in R (version 3.5.1) to assess the impact, if any, of invasive vegetation on male calling densities. Invasive vegetation features included in the analysis were: presence/absence of invasive species within the recording site, and the level (low/medium/high) of invasive vegetation adjacent to the recording site.

Results

Area searches for calling *A. rugosa* males yielded the discovery of 12 previously unknown *A. rugosa* populations. In total, calling individuals occupied 5.15 ha of land across the Klein Swartberg Mountain, representing 2.2% of the 230-ha suitable area. The acoustic surveys ($n = 7$ sites) covered over 50% (2.74 ha) of this area.

There was an average of $35,205 \pm 4,505$ detections of *A. rugosa* calls made during each 40-minute recording, of which a total of $16,962 \pm 1,740$ detections were included in the 2-minute subsamples ($n = 10$) used in the final analysis. The habitat patch area (in ha) and the mean number of calling males per ha (with their associated standard errors) for each site surveyed are displayed in Table 1. The mean advertisement call rate was 13.49 ± 0.97 calls per minute.

Across all survey sites, the total mean calling density was 417 males per ha (± 21.7 males per ha), and the estimated population size in the surveyed area (2.74 ha) was 1,053 males (± 194.1 with 95% CI). When considering both the survey area and the surrounding unsurveyed area (*i.e.*, the total occupied area; 5.15 ha), the estimated total population size was 2,060 males (± 132.2 with 95% CI). Therefore, assuming a male-to-female sex ratio of 1:1, the total adult population size is estimated to exceed 4,000 individuals.

Habitat assessments—The survey sites representing *A. rugosa* habitat were characterized by an average native vegetation height of 97.2 cm (± 8.2 cm; 95% CI) and 94.3% ($\pm 1.3\%$; 95% CI) ground cover. All seven survey sites were adjacent to streams that dispersed water throughout the seepage area, maintaining a moist environment. Sixteen *P. pinaster* saplings, with an average height of 46.8 cm (± 6.8 cm; 95% CI), were found within survey site 3 (western-most surveyed site within the eastern cluster of survey sites). No other surveyed areas were invaded by non-native vegetation, although much of the area surrounding surveyed sites within the western portion of the species' distribution (sites 1–3) was invaded by adult and juvenile *P. pinaster* and *H. sericea*. Significantly higher male calling densities were found at sites adjacent to low or no invasive vegetation, and no invasive species present in the surrounding habitat ($p < 0.0001$, $F_{3,211} = 23.82$, $R^2 = 0.2424$; Fig. 3).

Discussion

The *A. rugosa* total adult male population size was estimated to be 2,060 individuals (± 132.2 with 95% CI) using the newly developed acoustic spatially explicit capture-recapture (aSCR) technique. This estimate is more than five times higher than that formerly predicted by Turner and Channing (2008), where aural calling surveys projected the population to be 400 adults. At a sex ratio of 1:1, the total adult population is estimated to

Table 1. Results of vegetation assessments and the mean *Arthroleptella rugosa* calling male density per hectare and associated standard error (SE) for each replicate (n = 3) of all seven surveyed sites. The patch area (in hectares), mean ground cover (%), and mean vegetation height (cm) of each site is also reported.

Site	Rep	Mean calling male density (per ha)	Standard Error	Patch Area (ha)	Mean Ground Cover (%)	Mean Vegetation Height (cm)
1	1	465.18	45.79	0.20	96.8	185.5
	2	537.00	29.72			
	3	340.65	27.99			
2	1	334.87	25.17	0.24	90.5	98
	2	530.73	121.55			
	3	385.72	142.99			
3	1	360.54	33.20	0.44	89.0	126.5
	2	381.33	28.95			
	3	141.57	16.96			
4	1	456.78	27.26	0.79	94.1	54.0
	2	496.14	28.14			
	3	417.92	12.20			
5	1	790.51	21.91	0.15	96.0	67.1
	2	235.18	18.28			
	3	510.18	27.98			
6	1	181.54	12.85	0.36	98.5	84.9
	2	397.29	81.5			
	3	187.36	14.44			
7	1	169.08	25.50	0.56	95.0	58.9
	2	313.57	18.40			
	3	392.83	19.66			

exceed 4,000 individuals. This practical application of aSCR demonstrates how baseline population data can be determined for an entire species, providing population estimates that can be used in conservation assessments. Moreover, the methodology allows for repeated surveys with statistically robust estimates for monitoring purposes, where individual populations of the species can be targeted for repeated recordings (see Measey *et al.* 2017). These could be used to track the real impact of threats to species, such as invasive plants and fire (see Angus *et al.* 2023).

Arthroleptella rugosa populations are distributed within 2.2% of habitat patches across 230 ha of suitable habitat on the Klein Swartberg Mountain. Survey areas were, on average, characterized by high ground coverage and vegetation shorter than 1 m. Considering a large portion of the *A. rugosa* habitat was affected by fires in January 2012 (Measey *et al.* 2019), and fires have been associated with devastating adult frog population declines (Channing 2004, Measey *et al.* 2021), the estimates from this study suggest that the *A. rugosa* population has started to recover, even in densely invaded habitat where the destruction from fires would be most intense, (but see Angus *et*

al. [2023] for a description of the different recovery responses). However, it is likely that this apparent increase in population size was augmented by differences in the sampling techniques used and the additional populations discovered. The total population estimate from the present study was estimated by summing the population sizes for each surveyed and unsurveyed habitat patch, which covered the total patch area of 5.15 ha, even though the patches were not randomly sampled. In contrast, Turner and Channing (2008) conducted aural calling surveys within the six populations (2.78 ha) known at the time. The discovery, and subsequent inclusion, of the additional 2.37 ha of *A. rugosa* habitat in the present study would likely lead to higher population estimates. Alternatively, the currently implemented *P. pinaster* and *H. sericea* control across much of the Klein Swartberg Mountain could be adequately maintaining waterways, streams and seepage areas, sustaining suitable habitat for *A. rugosa* and facilitating steady population increases.

Although the total population appears to have increased from the previous estimation, the entire population of *A. rugosa* appears to be fragmented. A large *P. pinaster* stand in the

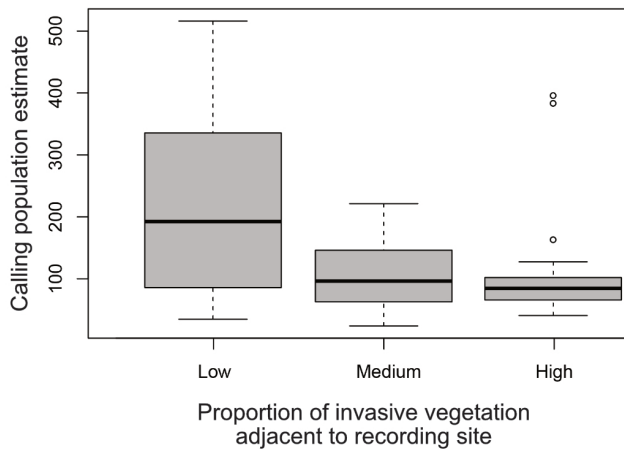


Fig. 3. The difference in calling densities of *Arthroleptella rugosa* from calling sites with different quantities of invasive pine and hakea in adjacent habitat in the Klein Swartberg, South Africa.

eastern and western clusters located in the middle of surveyed sites on the Klein Swartberg Mountain ridge (see Fig. 1; largest yellow polygon) could be acting as a barrier, physically and hydrologically separating the western populations from the remaining central and eastern populations. This would restrict the successful movement between and re-colonization of suitable habitat patches, affecting the dynamics of the meta-population and threatening *A. rugosa* persistence (Marsh and Trenham 2001). The hydrological distance between populations has been described as one of the most confounding factors affecting dispersal in small-sized mountain-dwelling anurans, as individuals tend to travel further along streams than through terrestrial spaces typically interspersed with unsuitable habitat (Measey *et al.* 2007). For direct-developing species like *A. rugosa*, where populations can occur continuously within habitats, extend past the ‘boundaries’ of suitable habitat areas and even occur throughout fragmented habitats (Measey *et al.* 2007), maintaining passive and active dispersal is integral to their conservation. As such, recognizing the importance of streams in maintaining connectivity across the *A. rugosa* distribution is essential to long-term population persistence.

Total population size and site occupancy of anurans can be highly variable, and so consistent, long-term monitoring of the *A. rugosa* population is necessary to assess population fluctuations and trends in relation to various conservation actions, environmental conditions, and other influential factors (Berven 1990). The effects of emerging invasive pines and hakea on *A. rugosa* populations might not be immediately apparent; however, the direct effects of fire would be (see Angus *et al.* 2023). This time lag may lead to difficulties in determining and targeting conservation efforts to mitigate all threatening processes affecting *A. rugosa*, as initially some threats may appear more severe than others (De Solla *et al.* 2006). Where possible, population survey information should be derived from long-term monitoring data.

The currently implemented management actions and higher than anticipated *A. rugosa* population size are evidence of the value of private reserve areas to amphibian conservation.

The Klein Swartberg Conservancy is a designated 14,857-ha area where both landowners and CapeNature officials have successfully implemented conservation actions directed at preserving *A. rugosa* populations and habitats. This conservancy provides the much-needed protection and restoration of not only *A. rugosa* habitat areas, but also the water systems that are critically endangered in the context of the broader landscape and the many other species that occur in this area. Thus, this conservancy is contributing directly to the conservation of a critically endangered species, and to the maintenance of waterways in the area. Monitoring of *A. rugosa* now requires the adoption and application of the aSCR method by CapeNature.

While the aSCR method accounts for distribution differences across the ESA, as estimates are averaged and extrapolated across all sites, this study assumes that the frogs are uniformly distributed across the occupied landscape. Population densities across the *A. rugosa* distribution, and thus also local population sizes, can vary significantly, as the landscape, altitude and habitat characteristics are not homogenous across the Klein Swartberg Mountain. This is evidenced within this study by the stark differences observed between the densities obtained for each of the surveyed sites (Table 1). Within survey areas, individuals may be unevenly distributed as they may associate with certain habitat features. Therefore, at both the survey area and full distribution scales, this assumption is likely to be violated. The present study attempted to address this through the novel incorporation of spatial population bounds, improving the precision of population estimates from each survey area. Furthermore, biases to the total population estimate would be mitigated through the inclusion of multiple survey areas to attain a representative sample of population densities across the *A. rugosa* distribution.

This method would be easily adoptable across various habitat types and species, as its implementation in the field does not require explicit user-training or expert species knowledge. It also has relatively low survey effort and can be adapted to a variety of spatial scales and to simultaneously monitor multiple species. Thus, aSCR presents the opportunity to implement monitoring to assess conservation efforts in both on and off reserve areas that have previously gone unchecked. Monitoring populations in these areas can identify the outcomes of implemented conservation actions, and thus audit their success. In this study, aSCR was used to rigorously estimate *A. rugosa* population densities, setting the precedent of using this technique on small, cryptic, vocally distinct species and providing the basis for commencing adequate monitoring schemes for many data deficient amphibian species. While this method is an effective conservation tool that can be used to maximize the efficiency of management actions, there are still many opportunities for further development. Advances in the incorporation of animal movement or individual animal recognition would significantly improve the precision of estimates and allow this method to be applied to any vocally distinct species. Furthermore, transitioning to a wireless, automated system would facilitate an easier collection of data without the presence of an observer. As aSCR methodologies are refined, this method will become an essential conservation tool used for monitoring acoustically

active species.

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Ethics statement

This study was carried out within the permission of the provincial conservation authority, CapeNature (Permit Number: AAA007-00162-0056) and was approved by the Animal Ethics Committee of the University of Queensland (Permit Number: ANRFA/SBS/280/15/STELLENBOSCH). Fieldwork was carried out on privately owned land, with access granted by landowners. The species targeted in acoustic surveys was a Critically Endangered species of frog, *Arthroleptella rugosa*. The survey methods chosen for this project were selected specifically to minimize researcher disturbance to the *A. rugosa* habitat, and extra care was taken in the field to ensure no unnecessary disturbance occurred.

Supplemental material

Original acoustic audio files: South African Environmental Observation Network (SAEON) Data Repository <https://catalogue.saeon.ac.za/records/10.15493/SAEON.SU.10000001>

Literature cited

- Angulo, A., M. Hoffmann, and G.J. Measey. 2011. Introduction: Conservation assessments of the amphibians of South Africa and the World. Ensuring a future for South Africa's frogs: A strategy for conservation research, p.1. SANBI Biodiversity Series 19. South African National Biodiversity Institute, Pretoria, South Africa.
- Angus, O., A.A. Turner, and J. Measey. 2023. In a rough spot: Declines in *Arthroleptella rugosa* calling densities are explained by invasive pine trees. *Austral Ecology* 48:498–512.
- Berven, K.A. 1990. Factors affecting population fluctuations in larval and adult stages of the Wood Frog (*Rana sylvatica*). *Ecology* 71:1599–1608.
- Borchers, D. 2012. A non-technical overview of spatially explicit capture–recapture models. *Journal of Ornithology* 152:435–444.
- Borchers, D. and R. Fewster. 2016. Spatial capture–recapture models. *Statistical Science* 31:219–232.
- Buckland, S.T., D.R. Anderson, K.P. Burnham, J.L. Laake, D.L. Borchers, and L. Thomas. 2001. Introduction to distance sampling: Estimating abundance of biological populations. Oxford University Press, Oxford, UK.
- Cardillo, M., G.M. Mace, J.L. Gittleman, K.E. Jones, J. Bielby, and A. Purvis. 2008. The predictability of extinction: Biological and external correlates of decline in mammals. *Proceedings of the Royal Society B* 275:1441–1448.
- Casazza, G., G. Paolo, B. Renato, F. Bruno, V. Daniele, F. Rossella, et al. 2014. Climate change hastens the urgency of conservation for range-restricted plant species in the central-northern Mediterranean region. *Biological Conservation* 179:129–138.
- Channing, A. 2004. Genus *Arthroleptella*. Pages 206–219 in L.R. Minter, M. Burger, J.A. Harrison, H.H. Braack, P.J. Bishop, and D. Kloepfer, editors. Atlas and red data book of the frogs of South Africa, Lesotho and Swaziland. Smithsonian Institution, Washington, DC, USA.
- Cooper, N., J. Bielby, G.H. Thomas, and A. Purvis. 2008. Macroecology and extinction risk correlates of frogs. *Global Ecology and Biogeography* 17:211–221.
- Cowling, R.M., D.M. Richardson, and P.J. Mustart. 1997. Fynbos. Pages 99–130 in R.M. Cowling, D.M. Richardson, and S.M. Pierce, editors. Vegetation of Southern Africa. Cambridge University Press, Cambridge, UK.
- Dawson, D.K. and M.G. Efford. 2009. Bird population density estimated from acoustic signals. *Journal of Applied Ecology* 46:1201–1209.
- De Solla, S., K. Fernie, G. Barrett, and C. Bishop. 2006. Population trends and calling phenology of anuran populations surveyed in Ontario estimated using acoustic surveys. Pages 113–129 in D. Hawksworth and A. Bull, editors. Marine, freshwater, and wetlands biodiversity conservation. Springer, Dordrecht, Netherlands.
- Dorcas, M.E., S.J. Price, S.C. Walls, and W.J. Barichivich. 2009. Auditory monitoring of anuran populations. Pages 281–298 in C.K. Dodd Jr., editor. Amphibian ecology and conservation: A handbook of techniques. Oxford University Press, New York, USA.
- Driscoll, D.A. 1998. Counts of calling males as estimates of population size in the endangered frogs *Geocrinia alba* and *G. vitellina*. *Journal of Herpetology* 32:475–481.
- Driver A., K.J. Sink, J.N. Nel, S. Holness, L. van Niekerk, F. Daniels, et al. 2012. National biodiversity assessment 2011: An assessment of South Africa's biodiversity and ecosystems. Synthesis Report. South African National Biodiversity Institute and Department of Environmental Affairs, Pretoria, South Africa.
- Efford, M. 2004. Density estimation in live-trapping studies. *Oikos* 106:598–610.
- Efford, M.G., D.K. Dawson, and D.L. Borchers. 2009. Population density estimated from locations of individuals on a passive detector array. *Ecology* 90:2676–2682.
- Gaston, K.J. and R.A. Fuller. 2009. The sizes of species' geographic ranges. *Journal of Applied Ecology* 46:1–9.
- Gillespie, D., D.K. Mellinger, J. Gordon, D. McLaren, P. Redmond, R. McHugh, et al. 2009. PAMGUARD: Semi-automated, open source software for real-time acoustic detection and localization of cetaceans. *Journal of the Acoustical Society of America* 125:2547.
- Harper, J.R.M., N.J. van Wilgen, A.A. Turner, K.A. Tolley, B. Maritz, S. Clusella-Trullas et al. 2022. Application of a trait-based climate change vulnerability assessment to determine management priorities at protected area scale. *Conservation Science and Practice* 4:e12756.
- International Union for Conservation of Nature and Natural Resources SSC Amphibian Specialist Group and South African Frog Re-assessment Group (IUCN and SA-FRoG). 2016. *Arthroleptella rugosa*. The IUCN Red List of Threatened Species 2016:



- e.T174664A77162276. Downloaded on 15 May 2017.
- Kohler, J., M. Jansen, A. Rodriguez, P. Kok, L. Toledo, M. Emmrich, *et al.* 2017. The use of bioacoustics in anuran taxonomy: Theory, terminology, methods and recommendations for best practice. *Zootaxa* 4251:1–124.
- Le Maitre, D.C., B.W. Van Wilgen, C.M. Gelderblom, C. Bailey, R.A. Chapman, and J.A. Nel. 2002. Invasive alien trees and water resources in South Africa: Case studies of the costs and benefits of management. *Forest Ecology and Management* 160:143–159.
- Lee, T.M. and W. Jetz. 2011. Unravelling the structure of species extinction risk for predictive conservation science. *Proceedings of the Royal Society B* 278:1329–1338.
- Marques, T.A., L. Thomas, S.W. Martin, D.K. Mellinger, S. Jarvis, R.P. Morrissey, *et al.* 2012. Spatially explicit capture–recapture methods to estimate minke whale density from data collected at bottom-mounted hydrophones. *Journal of Ornithology* 152:445–455.
- Marques, T.A., L. Thomas, S.W. Martin, D.K. Mellinger, J.A. Ward, D.J. Moretti, *et al.* 2013. Estimating animal population density using passive acoustics. *Biological Reviews* 88:287–309.
- Marsh, D.M. and P.C. Trenham. 2001. Metapopulation dynamics and amphibian conservation. *Conservation Biology* 15:40–49.
- Marsh, D.M., B.J. Cosentino, K.S. Jones, J.J. Apodaca, K.H. Beard, J.M. Bell, *et al.* 2017. Effects of roads and land use on frog distributions across spatial scales and regions in the Eastern and Central United States. *Diversity and Distributions* 23:158–170.
- Measey, G.J. 2011. Ensuring a future for South Africa’s frogs: a Strategy for conservation research. SANBI Biodiversity Series 19. South African National Biodiversity Institute, Pretoria, South Africa.
- Measey, G.J., P. Galbusera, P. Breyne, and E. Matthysen. 2007. Gene flow in a direct-developing, leaf litter frog between isolated mountains in the Taita Hills, Kenya. *Conservation Genetics* 8:1177–1188.
- Measey, G.J., B. Stevenson, T. Scott, R. Altwegg, and D. Borchers. 2017. Counting chirps: Acoustic monitoring of cryptic frogs. *Journal of Applied Ecology* 54:894–902.
- Measey, J., F. Becker and K.A. Tolley 2021. After the fire: Assessing the microhabitat of *Capensibufo rosei* (Hewitt, 1926). *Herpetology Notes* 14:169–175.
- Measey, J., J. Tarrant, A.D. Rebelo, A.A. Turner, L.H. Du Preez, M.M. Mokhatla, *et al.* 2019. Has strategic planning made a difference to amphibian conservation research in South Africa? *African Biodiversity & Conservation - Bothalia* 49:a2428.
- Meuche, I. and T.U. Grafe. 2005. Chorus tenure and estimates of population size of male European tree frogs *Hyla arborea*: Implications for conservation. *Amphibia-Reptilia* 26:437–444.
- Mokhatla, M.M., G.J. Measey, C.T. Chimimba, and B.J. van Rensburg. 2012. A biogeographical assessment of anthropogenic threats to areas where different frog breeding groups occur in South Africa: Implications for anuran conservation. *Diversity and Distributions* 18:470–480.
- Mucina, L., M.C. Rutherford, and L.W. Powrie editors. 2012. Vegetation map of South Africa, Lesotho and Swaziland (2012 beta). South African National Biodiversity Institute, Pretoria, South Africa.
- Nori, J., P. Lemes, N. Urbina-Cardona, D. Baldo, J. Lescano, and R. Loyola. 2015. Amphibian conservation, land-use changes and protected areas: A global overview. *Biological Conservation* 191:367–374.
- Parris K.M., T.W. Norton, and R.B. Cunningham. 1999. A comparison of techniques for sampling amphibians in the forests of south-east Queensland, Australia. *Herpetologica* 55:271–83.
- Pearson, R.G. 2006. Climate change and the migration capacity of species. *Trends in Ecology and Evolution* 21:111–113.
- Rebelo, A.D. and J. Measey. 2019. Locomotor performance constrained by morphology and habitat in a diverse clade of African frogs (Anura: Pyxicephalidae). *Biological Journal of the Linnean Society* 127:310–323.
- Ricketts, T.H., E. Dinerstein, T. Boucher, T.M. Brooks, S.H.M. Butchart, M. Hoffman, *et al.* 2005. Pinpointing and preventing imminent extinctions. *Proceedings of the National Academy of Sciences USA* 102:18497–18501.
- Rodrigues, A.S.L., S.J. Andelman, M.I. Bakarr, L. Boitani, T.M. Brooks, R.M. Cowling, *et al.* 2004. Effectiveness of the global protected area network in representing species diversity. *Nature* 428:640–643.
- Rodrigues, A.S.L., J.D. Pilgrim, J.F. Lamoreux, M. Hoffmann, and T.M. Brooks. 2006. The value of the IUCN Red List for conservation. *Trends in Ecology and Evolution* 21:71–76.
- Rogers, T.L., M.B. Ciaglia, H. Klinck, and C. Southwell. 2013. Density can be misleading for low-density species: Benefits of passive acoustic monitoring. *PLoS ONE* 8:e52542.
- Rouget, M., M. Barnett, R.M. Cowling, T. Cumming, F. Daniels, M.T. Hoffman, *et al.* 2014. Conserving the Cape Floristic Region. Pages 321–336 in N. Allsopp, J.F. Colville, and A. Verboom, editors. Fynbos: Ecology, evolution, and conservation of a megadiverse region. Oxford University Press, Oxford, UK.
- Sodhi, N.S., D. Bickford, A.C. Diesmos, T.M. Lee, L.P. Koh, B.W. Brook, *et al.* 2008. Measuring the meltdown: Drivers of global amphibian extinction and decline. *PLoS ONE* 3:e31636.
- Stevens, C.E., A.W. Diamond, and T.S. Gabor. 2002. Anuran call surveys on small wetlands in Prince Edward Island, Canada restored by dredging of sediments. *Wetlands* 22:90–99.
- Stevenson, B.C. and D.L. Borchers. 2015. SECR models with supplementary location information. <https://github.com/b-steve/admbsec>. R package version 1.1.0. Downloaded on 15 May 2017.
- Stevenson, B.C., D.L. Borchers, R. Altwegg, R.J. Swift, D.M. Gillespie, and G.J. Measey. 2015. A general framework for animal density estimation from acoustic detections across a fixed microphone array. *Methods in Ecology and Evolution* 6:38–48.
- Stevenson, B., P. van Dam-Bates, C.K.Y. Young, and J. Measey. 2021. A spatial capture-recapture model to estimate call rate and population density from passive acoustic surveys. *Methods in Ecology and Evolution* 12:432–442.
- Stuart, S.N., J.S. Chanson, N.A. Cox, B.E. Young, A.S.L. Rodrigues, D.L. Fischman, *et al.* 2004. Status and trends of amphibian declines and extinctions worldwide. *Science* 306:1783–1786.
- Turner, A.A., editor. 2012. Western Cape Province state of biodiversity 2012. CapeNature Scientific Services, Stellenbosch, South Africa.
- Turner, A. and A. Channing. 2008. A new species of *Arthroleptella* Hewitt, 1926 (Anura: Pyxicephalidae) from the Klein Swartberg

- Mountain, Caledon, South Africa. *African Journal of Herpetology* 57:1–12.
- Turner, A. and A. Channing. 2017. Three new species of *Arthroleptella* Hewitt, 1926 (Anura: Pyxicephalidae) from the Cape Fold Mountains, South Africa. *African Journal of Herpetology* 66:53–78.
- Tyre, A.J., B. Tenhumberg, S.A. Field, D. Niejalke, K. Parris, and H.P. Possingham. 2003. Improving precision and reducing bias in biological surveys: Estimating false-negative error rates. *Ecological Applications* 13:1790–801.
- Van der Meijden, A., A. Crottini, J. Tarrant, A. Turner, and M. Vences. 2011. Multi-locus phylogeny and evolution of reproductive modes in the Pyxicephalidae, an African endemic clade of frogs. *African Journal of Herpetology* 60:1–12.
- Venter, O., R.A. Fuller, D.B. Segan, J. Carwardine, T. Brooks, S.H.M. Butchart, *et al.* 2014. Targeting global protected area expansion for imperiled biodiversity. *PLoS Biology* 12:e1001891.
- Wilson, J.R., M. Gaertner, C.L. Griffiths, I. Kotze, D.C. Le Maitre, S.M. Marr, *et al.* 2014. Biological invasions in the Cape Floristic Region: History, current patterns, impacts, and management challenges. Pages 273–298 in N. Allsopp, J.F. Colvill, and A. Verboom, editors. *Fynbos: Ecology, evolution, and conservation of a megadiverse region*. Oxford University Press, Oxford, UK.
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Stark, D., *et al.* 2024. Population estimation of a cryptic moss frog using acoustic spatially explicit capture recapture. Pages 110–119 in S.C. Walls and K.M. O'Donnell, editors. *Strategies for Conservation Success in Herpetology*. Society for the Study of Amphibians and Reptiles, University Heights, OH, USA.